



AFLATOXINS IN FOOD GRAINS: CONTAMINATION, DANGERS AND CONTROL

*Abdullahi, N. and Dandago, M. A.

Department of Food Science and Technology, Kano University of Science and Technology, Wudil P. M. B. 3244 Kano, Kano State, Nigeria.

*Corresponding Author's email: nurafst@gmail.com

ABSTRACT

The most concern postharvest safety issue in food grains is aflatoxins production in contaminated stored grains. Consumption of aflatoxins contaminated grains can lead to complicated health issues that can lead to death. Aflatoxins are secondary metabolites commonly produced by *Aspergillus flavus* and *Aspergillus parasiticus*. They were reported to disturb foetus development, causes changes in living cells, suppresses the immune system and causes many forms of cancers. Aflatoxin B₁ was classified under group I carcinogens by the International Agency for Research on Cancer. Aflatoxins contamination affects food security and can hinder international trade due to the strict ban enforce by many nations. Contaminations were reported in raw and processed grains (including ready-to-eat), milk and meat of farm animals and human breastmilk and blood. Major causes of grains aflatoxins contamination are wrong harvesting time and method, improper drying, poor storage and processing and higher moisture in the grains. Literature was gathered through an online search on Google Scholar, attention was given to the articles published in the last 5 years. Causes of fungal contamination, aflatoxins production and their control measures were deliberated, possible means of mitigating aflatoxins contamination through consumption of food grains were also recommended.

Keywords: mycotoxins, storage, drying, cereals, legumes

INTRODUCTION

Fungi can attack grains before or after drying and cause serious damage to raw materials and processed foods during storage and transportation (Aldars-García *et al.*, 2016). Appropriate drying to a moisture content below microbial thriving levels is critical to the stability, postharvest storage and processing qualities of all stable grains (Bradford *et al.*, 2018). Inappropriate storage and packaging materials that allow moisture permeation rises relative humidity during storage and favours insects and microbial activities (Bradford *et al.*, 2018). Postharvest grain losses cause great constrain to food and nutritional security (Affognon *et al.*, 2015; Likhayo *et al.*, 2018; Mezgebe *et al.*, 2016), economy and health (Khaneghah *et al.*, 2018; Kumari *et al.*, 2020).

Aflatoxins (AFs) are immunosuppressive, carcinogenic (Kachapulula *et al.*, 2017), mutagenic and teratogenic (Blankson and Mill-Robertson, 2016; Kebede *et al.*, 2020) secondary metabolites produced by fungi (Aron *et al.*, 2017), mainly *Aspergillus* (Eljack, 2012; Maringe *et al.*, 2016; Valencia-Quintana *et al.*, 2020), *Fusarium* (Kebede *et al.*, 2020) and *Penicillium* (Elias, 2016) during storage of contaminated grains. *Aspergillus flavus* and *Aspergillus parasiticus* occurs more frequently than other fungi in grains and produced more AFs including AFB₁ (Mom *et al.*, 2020). Ezekiel *et al.* (2020) recently discovered new fungi species suspected to be potential toxin producers. Mycotoxins production depends on certain environmental factors, fungi grow optimally in a humid environment rich in nutrients essential to them (Adeyeye, 2016). Grains can be contaminated by either field fungi, store fungi, or both (Ncube and Maphosa, 2020; Tola and Kebede, 2016). Ramirez *et al.* (2018) reported changes in the population and types of fungi during the storage

of chickpea. AFs contamination accounts for about 25 % of global crop loss (Serdar *et al.*, 2020).

There are four types of AFs; B₁, B₂, G₁ and G₂ (Ahmadi *et al.*, 2020). Aflatoxin B₁ (AFB₁) is the most mutagenic, carcinogenic and teratogenic material found in foods (Ahmadi *et al.*, 2020; Nesci *et al.*, 2016). It is the most potent known liver carcinogen (Jallow *et al.*, 2018). It was classified under group I carcinogens by the International Agency for Research on Cancer (Al-Zoreky and Saleh, 2019). AFB₁ is very stable and can be toxic at a meagre dose (Ponzilacqua *et al.*, 2018). The menace of AFB₁ is common to many staple foods in most developing countries. Groundnut is the most disposed crop to AFB₁ contamination (Jallow *et al.*, 2018). More than 90 % of food samples collected by Eshete *et al.* (2020) from Sidama zone, Ethiopia contains AFB₁ above the EU permissible limit. Traditionally processed infant food in Ouagadougou, Burkina Faso contains AFB₁ 900 times higher than the EU limit of 0.1 µg/kg (Ware *et al.*, 2017). More than 41 % of maize market samples in Ghana contain AFB₁ above Ghana and EU permissible limits (Kortei *et al.*, 2021). Sulaiman *et al.* (2018) associated urine AFB₁ with cereal products consumption. AFB₁ estimated daily intake of 0.23 µg/kg/bwd and 0.153 µg/kg/bwd were reported in Ghanaian infants and children respectively (Blankson and Mill-Robertson, 2016). The results of a laboratory experiment conducted by Branà *et al.* (2017) showed that *Plerotus eryngii* (king oyster mushroom) can degrade AFB₁ in malt extract broth.

In addition to AFs, fungi also produced other carcinogenic and mutagenic secondary metabolites such as cyclopiazonic acid, aflatrem (Ojiambo *et al.*, 2018), ochratoxin A, fumonisins (Sun *et al.*, 2017), deoxynivalenol, zearalenone, T-2 toxin and HT-2 toxin (Kunz *et al.*, 2020). *Fusarium graminearum* produces

zearalenone and deoxynivalenol in grains particularly during slow drying or due to higher moisture in stored grains (Portell *et al.*, 2020). Ochratoxin A and fumonisins were reported to cause renal cancer and liver cancer respectively (Huong *et al.*, 2016).

The menace of mycotoxins contamination is a threat to food safety during storage and distribution (Aldars-García *et al.*, 2016). AFs are persistent and their contamination can occur before or during harvesting (Kortei *et al.*, 2019) and at various points along the supply chain depending on the handling condition (Mannaa and Kim, 2017; Neme and Mohammed, 2017; Udomkun *et al.*, 2017). Bamba *et al.* (2021) reported AFs contamination in the spathes, cobs and grains of maize. Contamination occurs mostly during drying and storage (Njoroge *et al.*, 2019; Quellhorst *et al.*, 2020).

Aflatoxins Contamination

Postharvest AFs contamination occurs when aflatoxigenic strains of *Aspergillus* contaminates grains before harvest (Waliyar *et al.*, 2015). Global consumption of AFs ranged from 3.0 to 17.1 ng/kg bw/day with rice, wheat and maize having the highest contributions (Andrade and Caldas, 2015). The maximum permissible limit for AFs in grains is 10 µg/kg (10 ppb) (Ayelign *et al.*, 2018).

AFs contamination occurs in grains due to exposure to certain pre-and post-harvest conditions (Aron *et al.*, 2017). Contaminations are mostly associated with poor agricultural practices (Nleya *et al.*, 2018). Tuan *et al.* (2018) reported that non-dormant seeds and that with low levels of dormancy can germinate while attached to the mother plant, this can reduce yield and trigger mould growth and AFs accumulation before harvest. Kachapulula *et al.* (2017) reported that uncultivated lands are loaded with more AFs producing organisms than cultivated lands. Mould growth and AFs contamination depend greatly on grain moisture content and temperature and relative humidity of the storage or packaging environment (Neme and Mohammed, 2017; Tola and Kebede, 2016).

Major factors that account for AFs contamination are wrong harvesting time and method, improper drying, inadequate storage and processing and higher moisture in the grains (Chukwukere *et al.*, 2021; Neme and Mohammed, 2017). Aron *et al.* (2017) also reported that poor postharvest handling of grains causes AFs contamination. Seetha *et al.* (2017) witnessed AFB₁ increase in maize, sorghum, Bambara nut, groundnut, and sunflower during storage. Tibagonzeka *et al.* (2018) reported that grains dried on the bare surface are more prone to AFs contamination than grains dried on a covered surface.

Except for few crops such as rice, many kinds of cereal and legumes are disposed to AFs contamination (Gonçalves *et al.*, 2019; Sun *et al.*, 2017). Contaminations were reported during the pre-and postharvest stages in groundnuts, millet, sesame seeds, maize, wheat, rice, fig, spices, cocoa, and processed products such as peanut butter, cooking oil (Mahato *et al.*, 2019), bakery products, coffee, macaroni (Serdar *et al.*, 2020) and processed flours (Adeyeye, 2016) among other grain products. Legumes in general are more prone to AFs contamination (Eljack, 2012) and groundnut is more disposed than other legumes (Maringe *et al.*, 2016). Sorghum, groundnut and maize contributed to AFs contamination more than other grains (Neme and Mohammed, 2017; Tibagonzeka *et al.*, 2018), their prevalence of AFs contamination can be as high as 91 %, 55 % and 44 % respectively (Tibagonzeka *et al.*, 2018). Maize, peanut and peanut oil are the most disposed food to AFs

contamination in sub-Sahara Africa (Ingenbleek *et al.*, 2019). Higher temperatures and relative humidity facilitate mould growth and AFs formation (Sun *et al.*, 2017; Valencia-Quintana *et al.*, 2020). The prevailing weather condition in sub-Sahara Africa characterized by heavy downfall and higher temperatures, facilitate mould growth and AFs production in grains (Ingenbleek *et al.*, 2019; Ncube and Maphosa, 2020; Nleya *et al.*, 2018).

The danger is more common and likewise more extreme in developing countries (Udomkun *et al.*, 2017). Mycotoxins contamination is more common in African countries due to poor socio-economic conditions (Kebede *et al.*, 2020). The results of the survey conducted by Wangia-Dixon *et al.* (2020) in Makueni and Siaya Counties, Kenya showed that children from low-income families are more prone to AFs contamination. About 49 % of complimentary food samples collected by Aron *et al.* (2017) from Bahi District, Tanzania reported being contaminated with AFs. A significant proportion of maize, sorghum and millet samples collected from Makueni and Nandi, Kenya contains AFs and fumonisins above the recommended levels of 10 ppb and 2 ppm respectively (Kang'Ethe *et al.*, 2017). The AFs levels in recently harvested groundnuts, beans, cowpeas and Bambara nuts samples from Shamva and Makoni districts, Zimbabwe is alarming and may significantly upsurge during storage (Maringe *et al.*, 2016). Blankson *et al.* (2019) reported that 96 % of processed infant food sold in Accra, Ghana possessed AFs above the EU permissible limit. The finding of Obade (2015) revealed that weaning foods commonly used Kisumu County, Kenya are contaminated with AFs above permissible amounts. About 93 and 42 % of household and industrially processed foods samples respectively collected from Lagos and Ogun States, Nigeria are contaminated with mycotoxins including AFs (Ojuri *et al.*, 2019). Rice and beans samples collected from Enugu, Nigeria contain AFs above permissible limits (Dozie-Nwakile *et al.*, 2020). Likewise, maize samples collected from Dutsinma, Nigeria was heavily contaminated with *Aspergillus* fungi (Mzungu *et al.*, 2018). About 58 % of basmati rice samples collected from Lahore, Narowal, Faisalabad and Multan, Pakistan reported to content aflatoxins above permissible limits (Mukhtar *et al.*, 2016). Aflatoxins content between 0.09 and 579 µg/kg were reported in nutty food samples collected from Jidda markets in Saudi Arabia (Tawila *et al.*, 2020). More than 95 % of complimentary food samples collected from Amhara, Tigray and Oromia, Ethiopia were contaminated with AFs (Ayelign *et al.*, 2018). About 72 % of infant foods samples collected from Accra, Ghana contains AFB₁ above EU permissible limits (Blankson and Mill-Robertson, 2016). Total AFs content in maize, sorghum and millet flours collected from commercial milling centers in Nairobi, Kenya was found to be 59.73, 39.21 and 34.80 µg/kg respectively (Wanjeri *et al.*, 2017). Eshete *et al.* (2020) detected AFs above EU permissible limits in 5.3 % of breastmilk samples collected from Sidama zone, Ethiopia.

Developing countries with strict food regulations reported lower levels of AFs contamination. Values within permissible limits were reported by Ahmadi *et al.* (2020) in peas, red beans, lentils, mung bean and cotyledons samples collected from Tehran, Iran. Likewise, rice samples collected from imported bulk in Saudi Arabia (Al-Zoreky and Saleh, 2019).

Fungal hydrolytic enzymes also lead to the degradation of proteins and carbohydrates in grains, this can lead to a poor

quality dough and bad baked products (Schmidt, 2018). AFs contamination is seriously affecting the economy of many developing nations due to strict AFs contamination regulations in international trade (Eljack, 2012; Tumukunde *et al.*, 2020; Udomkun *et al.*, 2017).

Dangers Associated with Aflatoxins Contamination in Grains

Inappropriate handling and storage of grains after harvesting expose them to optimum conditions for fungal growth and AFs production (Waliyar *et al.*, 2015). The quality and safety of processed grains and their by-products depend to the large extent on the pre- and post-harvest qualities of the grains (Pratap *et al.*, 2016). The danger of microbial postharvest losses is beyond losing grains qualities, it can as well lead to dangerous health problems including cancers (Schmidt *et al.*, 2018). Mycotoxin (including AFs) contaminations through consumption of inadequately dried and contaminated foods are affecting about 4.5 billion people in the world (Bradford *et al.*, 2018). AFs can pass through the metabolic system unchanged and accumulate in body organs (Khaneghah *et al.*, 2018). Therefore, AFs can be found in the milk and meat of farm animals fed on the contaminated diets (Wangia-Dixon *et al.*, 2020). AFs were also found in human serum (Sabuncuoglu *et al.*, 2015) and breastmilk (Eshete *et al.*, 2020).

Consumption of foods contaminated with AFs is a threat to human and animal health (Mannaa and Kim, 2017; Ojiambo *et al.*, 2018). Depending on cereal legume-based composites exclusively as complimentary food exposed children to malnutrition and AFs and fumonisins contaminations (Mollay *et al.*, 2021). AFs contamination can lead to poor growth and development (Achaglinkame *et al.*, 2017), vaccine interference (Wangia-Dixon *et al.*, 2020) and iron deficiency (Opoku *et al.*, 2018) in infants and children. Can also lead to grave health issues including liver cancer (Maringe *et al.*, 2016), immune suppression, embryo toxicity and nutritional deficiencies (Granados-Chinchilla *et al.*, 2017). Acute aflatoxicosis can lead to haemorrhage, severe liver damage, oedema, and death (Khaneghah *et al.*, 2018). In addition to aflatoxicosis, food fungal contamination can also cause other serious diseases such as aspergilloma (Dozie-Nwakile *et al.*, 2020) and infections in patients with immune-compromised and hypersensitive reactions such as asthma and allergic alveolitis (Muhie and Bayisa, 2020).

Control of Aflatoxins Contamination in Grains

Controlling AFs in grains is quite challenging because many fungal species are toxigenic and their mycotoxins synthetic pathways and factors affecting them are not yet fully understood (Aldars-García *et al.*, 2016). Postharvest decontamination of grains is crucial to the postharvest quality and safety of grains and their products (Schmidt *et al.*, 2019). Preventing the growth of AFs producing microorganisms will inevitably prevent AFs contamination in cereals and legumes (Achaglinkame *et al.*, 2017). Sirma *et al.* (2018) opined that the addition of AFs absorbents and enzymes can significantly lower AFs contamination. Many plant extracts and essential oils were reported to mitigate fungal growth and AFs production (Ponzilacqua *et al.*, 2018). Telles *et al.* (2017) reported that peanut and azuki bean phenolic extracts can protect beans against fungal contamination and AFs production.

AFs contamination can effectively be control in grains through decent agricultural practice, (Achaglinkame *et al.*, 2017; Gonçalves *et al.*, 2019), production of fungi resistant varieties

(Ncube and Maphosa, 2020), fast and proper drying, insects control, use of natural and synthetic antifungal, irradiation (Neme and Mohammed, 2017), cleaning and sanitizing storage facilities, avoiding conditions that will favour mold growth and AFs production (Gagiu *et al.*, 2018), avoiding grains damage, ensuring good postharvest practices (Mannaa and Kim, 2017), prevention legislation and policy (Khaneghah *et al.*, 2018) and public enlightenment (Achaglinkame *et al.*, 2017; Michael *et al.*, 2018). Training farmers on AFs contamination mitigating techniques yielded positive results in Tanzania (Seetha *et al.*, 2017). Nesci *et al.* (2016) recommended the use of food-grade antioxidants microcapsules in the prevention of fungal attacks and AFs production. Incorporation of savannah tea leaves as bio-pesticide into hermetic bag reduces aflatoxins contamination in cowpea during 8 months of storage (Constant *et al.*, 2016).

Cleaning and processing operations such as sorting, milling, fermentation, roasting, baking, flaking and extrusion cooking are reported to lower mycotoxins levels in food (Neme and Mohammed, 2017). Products of lactic acid fermentation were reported to prevent AFs synthesis (J. Prakash, 2016). Ibitoye *et al.* (2020) reported a decrease in the growth rate of *A. flavus* and a reduction in AFB₁ and AFB₂ production in sorghum and millet treated with monoculture and co-culture LAB. Processing methods and severity also affect AFs concentration, Ojuri *et al.* (2019) reported that household processed foods are more contaminated with mycotoxins than industrially proceed foods. Kamala *et al.* (2018) reported that sorting by handpicking, proper sun drying on an elevated surface, chemical treatment before storage and dehulling of maize before milling lower AFs intake in Tanzanian infants by 78 %. Microwaving, vacuum packaging and high hydrostatic pressure inhibited fungal growth and AFs production in wheat (Schmidt *et al.*, 2019). Waliyar *et al.* (2015) reported a higher concentration of AFB₁ in groundnut paste than in groundnut seed in market samples collected from Kolokani, Kayes, and Kita districts, Mali. The higher concentration of the AF in the processed product may result from post-process exposure to environmental conditions that favours AFs production. The finding of Udomkun *et al.* (2019) showed that combining staple grains with other locally available crops reduces grains AFs contamination. While combining other grains with groundnut drastically increase AFs in the resultant composite (Temba *et al.*, 2017). Similarly, much higher contaminations were reported in cereal legume-based foods than in cereal-based products (Opoku *et al.*, 2018).

A meaningful advancement in controlling AFs contamination was reported in biological control using atoxigenic (nonAFs producing) strains of *A. flavus* (Ojiambo *et al.*, 2018). Sarrocco and Vannacci (2018) reviewed the possibility of applying beneficial fungi at the pre-harvest stage to prevent postharvest fungal contamination and mycotoxins production in cereals, apples and grapes. N₂ saturated atmosphere (98.5 %) greatly reduces growth and sporulation of all AFs producing *Aspergillus* and destroy *Sitophilus oryzae* and *Tribolium confusum* after 3 and 7 days in wheat and its flour respectively (Lorenzo *et al.*, 2020).

AFs contamination in developing countries requires a collective approach that will simultaneously consider food safety, food production and humans and animal health (Aron *et al.*, 2017). Most consumers in developing countries are not familiar with the dangers associated with the consumption of moldy foods

(Adeyeye, 2020). Njoroge (2018) opined that success in the fight against AFs contamination will only be achieved in Africa if consumers realised the dangers associated with the consumption of AF-contaminated foods and start demanding better quality and safe foods. Technical and financial support from international donors is required to minimise or eliminate AFs contamination since many developing countries lack adequate resources (Elias, 2016). Achaglinkame *et al.* (2017) recommended the substitution of cereal-legume blend with a tuber-based blend for infant formula, this could not provide some essential nutrients and can only be possible if the blends will be enriched with the essential nutrients deficient in tuber crops. Farmers and other stakeholders in Africa need to be sensitized on the activities of Partnership for Aflatoxin Control in Africa (PACA), stakeholders are required to present valid evidence for AFs contamination to access AFs mitigation technologies (Njoroge, 2018).

Recommendations

1. Consumers should avoid crops with higher AFs accumulation. Sorghum, groundnut and maize were reported to habitually accumulate AFs. These can be substitute by underutilized grains such as millet which was reported to have excellent nutritional and health benefits (Birania *et al.*, 2020)
2. Recently harvested grains with alarming levels of fungi and/or AFs contaminations should be processed immediately to avoid AFs accumulation during storage
3. Countries in sub-Sahara Africa need to urgently develop and enact regulations on AFs food contamination as the region is the most reported in the literature, possibly due to the prevailing weather condition that favours fungal growth.
4. Researches in plant genetics and molecular biology should be focused on developing grains that will be resistant to fungal contamination. The development of insect and mold resistance grains will surely improve safety and minimize storage challenges particularly among farmers that cannot afford modern storage and packaging materials.
5. The possibility of eliminating *Aspergillus* fungus during the storage of grains through microbiological succession using benign microorganisms should be studied.
6. Microorganisms and insects continue to develop resistance to the existing various postharvest treatments, hence unceasing research is necessary in this area.

CONCLUSION

The jeopardy of AFs contamination is a threat to public health in many developing countries. Inappropriate storage and packing systems with higher humidity and temperature allow the growth and proliferation of aflatoxins producing fungi. Contamination can also occur during harvest and pre-harvest time. Fungi metabolites can contaminate up to 25 % of stored grains when exposed to unsuitable storage conditions. Higher levels of contaminations were reported in legumes grains, however, staples grains such as wheat, rice and maize account for greater contaminations in humans. Control of AFs contamination requires a holistic approach that will protect throughout the supply chain. The danger can be mitigated by

decent agricultural, storage and processing practices; including production of resistance varieties, proper and adequate drying, insect and mold control during storage, good manufacturing practices including proper and adequate cleaning and processing, education and enlightenment, and extenuating legislation and policy that will ensure acceptance of only good quality commodities.

REFERENCES

- Achaglinkame, A. M., Opoku, N., & Amagloh, F. K. (2017). Aflatoxin contamination in cereals and legumes to reconsider usage as complementary food ingredients for Ghanaian infants: A review. *Journal of Nutrition and Intermediary Metabolism*, *10*, 1–7. <https://doi.org/10.1016/j.jnim.2017.09.001>
- Adeyeye, S. A. O. (2016). Fungal mycotoxins in foods: A review. *Cogent Food & Agriculture*, *2*(1), 1–11. <https://doi.org/10.1080/23311932.2016.1213127>
- Adeyeye, S. A. O. (2020). Aflatoxigenic fungi and mycotoxins in food: a review. *Critical Reviews in Food Science and Nutrition*, *60*(5), 709–721. <https://doi.org/10.1080/10408398.2018.1548429>
- Affognon, H., Mutungi, C., Sanginga, P., & Borgemeister, C. (2015). Unpacking postharvest losses in sub-Saharan Africa: A Meta-Analysis. *World Development*, *66*, 49–68. <https://doi.org/10.1016/j.worlddev.2014.08.002>
- Ahmadi, M., Jahed Khaniki, G., Shariatifar, N., & Molae-Aghaee, E. (2020). Investigation of aflatoxins level in some packaged and bulk legumes collected from Tehran market of Iran. *International Journal of Environmental Analytical Chemistry*, *1*–10. <https://doi.org/10.1080/03067319.2020.1789614>
- Al-Zoreky, N. S., & Saleh, F. A. (2019). Limited survey on aflatoxin contamination in rice. *Saudi Journal of Biological Sciences, In Press*. <https://doi.org/10.1016/j.sjbs.2017.05.010>
- Aldars-García, L., Ramos, A. J., Sanchis, V., & Marín, S. (2016). Modeling postharvest mycotoxins in foods: recent research. *Current Opinion in Food Science*, *11*, 46–50. <https://doi.org/10.1016/j.cofs.2016.09.005>
- Andrade, P. D., & Caldas, E. D. (2015). Aflatoxins in cereals: Worldwide occurrence and dietary risk assessment. *World Mycotoxin Journal*, *8*(4), 415–431. <https://doi.org/10.3920/WMJ2014.1847>
- Aron, L., Makangara, J. J., Kassim, N., & Ngoma, S. J. (2017). Post-harvest Practices Associated with Aflatoxins Contamination of Complementary Flours in Bahi District, Dodoma, Tanzania. *International Journal of Sciences: Basic and Applied Research*, *36*(6), 174–186. <http://gssrr.org/index.php?journal=JournalOfBasicAndApplied>
- Ayalign, A., Woldegiorgis, A. Z., Adish, A., & De Saeger, S. (2018). Total aflatoxins in complementary foods produced at community levels using locally available ingredients in Ethiopia. *Food Additives and Contaminants: Part B Surveillance*, *11*(2), 111–118. <https://doi.org/10.1080/19393210.2018.1437784>

- Bamba, S., Biego, H. M. G., Coulibaly, A., Yves, N. B., & Daouda, S. (2021). Determination of the Level of Aflatoxins Contamination in Maize (*Zea mays* L.) Produced in Five Regions of Côte d'Ivoire. *Asian Research Journal of Agriculture*, 14(2), 21–31. <https://doi.org/10.9734/arja/2021/v14i230121>
- Birania, S., Rohilla, P., Kumar, R., & Kumar, N. (2020). Post harvest processing of millets: A review on value added products. *International Journal of Chemical Studies*, 8(1), 1824–1829. <https://doi.org/10.22271/chemi.2020.v8.i1aa.8528>
- Blankson, G. K., & Mill-Robertson, F. C. (2016). Aflatoxin contamination and exposure in processed cereal-based complementary foods for infants and young children in greater Accra, Ghana. *Food Control*, 64, 212–217. <https://doi.org/10.1016/j.foodcont.2015.12.032>
- Blankson, G. K., Mills-Robertson, F. C., & Ofori, I. W. (2019). Survey of occurrence levels of Aflatoxins in selected locally processed cereal-based foods for human consumption from Ghana. *Food Control*, 95, 170–175. <https://doi.org/10.1016/j.foodcont.2018.08.005>
- Bradford, K. J., Dahal, P., Van Asbrouck, J., Kunusoth, K., Bello, P., Thompson, J., & Wu, F. (2018). The dry chain: Reducing postharvest losses and improving food safety in humid climates. *Trends in Food Science and Technology*, 71, 84–93. <https://doi.org/10.1016/j.tifs.2017.11.002>
- Branà, M. T., Cimmarusti, M. T., Haidukowski, M., Logrieco, A. F., & Altomare, C. (2017). Bioremediation of aflatoxin B1-contaminated maize by king oyster mushroom (*Pleurotus eryngii*). *PLoS ONE*, 12(8), 1–14. <https://doi.org/10.1371/journal.pone.0182574>
- Chukwukere, V., Amah, N., & Jabil, I. (2021). Perceived Causes of Aflatoxin Contamination of Cereal and Legume Grains on Rural Farmers' Livelihood In Jos South Local. *International Journal of Science and Applied Research*, 4(1), 39–44.
- Constant, K. K., Adama, C., Daouda, S., Olivier, C., Godi, B., & Marius, H. (2016). Evolution of Aflatoxins Levels during Storage of Cowpeas (*Vigna unguiculata* L Walp) Bagged Pics Containing *Lippia multiflora* Moldenke Leaves and Ivorian Exposure Risk. *International Journal of Science and Research (IJSR)*, 5(7), 678–691. <https://doi.org/10.21275/v5i7.art2016285>
- Dozie-Nwakile, O., Onyemelukwe, N., Nwakile, C., Okenwa, C., Okongwu, U., Ukpai, N., & Ilo, A. (2020). Nutritional Sustainability for a Child towards Isolation of *Aspergillus* Species from Some Cereals and Legumes Sold in Enugu. *Acta Scientifica Nutritional Health*, 4(5), 55–59. <https://doi.org/10.31080/asnh.2020.04.0702>
- Elias, N. K. S. (2016). Aflatoxins: A silent threat in developing countries. *African Journal of Biotechnology*, 15(35), 1864–1870. <https://doi.org/10.5897/ajb2016.15305>
- Eljack, A. E. T. M. (2012). *Level of Contamination with the Fungus (Aspergillus flavus) and Aflatoxins in Some Legume seeds and Cereal Grains*. University of Gezira.
- Eshete, M., Gebremedhin, S., Alemayehu, F. R., Taye, M., Boshe, B., & Stoecker, B. J. (2020). Aflatoxin contamination of human breast milk and complementary foods in southern Ethiopia. *Maternal and Child Nutrition*, 17(e13081). <https://doi.org/10.1111/mcn.13081>
- Ezekiel, C. N., Kraak, B., Sandoval-Denis, M., Sulyok, M., Oyedele, O. A., Ayeni, K. I., Makinde, O. M., Akinyemi, O. M., Krska, R., Crous, P. W., & Houbaken, J. (2020). Diversity and toxigenicity of fungi and description of *Fusarium madaense* sp. nov. From cereals, legumes and soils in north-central Nigeria. *MycKeys*, 67, 95–124. <https://doi.org/10.3897/MYCOKEYS.67.52716>
- Gagiu, V., Mateescu, E., Armeanu, I., Dobre, A. A., Smeu, I., Cucu, M. E., Oprea, O. A., Iorga, E., & Belc, N. (2018). Post-harvest contamination with mycotoxins in the context of the geographic and agroclimatic conditions in Romania. *Toxins*, 10, 1–17. <https://doi.org/10.3390/toxins10120533>
- Gonçalves, A., Gkrillas, A., Dorne, J. L., Dall'Asta, C., Palumbo, R., Lima, N., Battilani, P., Venâncio, A., & Giorni, P. (2019). Pre- and Postharvest Strategies to Minimize Mycotoxin Contamination in the Rice Food Chain. *Comprehensive Reviews in Food Science and Food Safety*, 18(2), 441–454. <https://doi.org/10.1111/1541-4337.12420>
- Granados-Chinchilla, F., Molina, A., Chavarría, G., Alfaro-Cascante, M., Bogantes-Ledezma, D., & Murillo-Williams, A. (2017). Aflatoxins occurrence through the food chain in Costa Rica: Applying the One Health approach to mycotoxin surveillance. *Food Control*, 82, 217–226. <https://doi.org/10.1016/j.foodcont.2017.06.023>
- Huong, B. T. M., Tuyen, L. D., Tuan, D. H., Brimer, L., & Dalsgaard, A. (2016). Dietary exposure to aflatoxin B1, ochratoxin A and fumonisins of adults in Lao Cai province, Viet Nam: A total dietary study approach. *Food and Chemical Toxicology*, 98, 127–133. <https://doi.org/10.1016/j.fct.2016.10.012>
- Ibitoye, O. A., Olaniyi, O. O., Ogidi, C. O., & Akinyele, B. J. (2020). Lactic acid bacteria bio-detoxified aflatoxins contaminated cereals, ameliorate toxicological effects and improve haemato-histological parameters in albino rats. *Toxin Reviews*, 1–12. <https://doi.org/10.1080/15569543.2020.1817088>
- Ingenbleek, L., Sulyok, M., Adegboye, A., Hossou, S. E., Koné, A. Z., Oyedele, A. D., Kisito, C. S. K. J., Dembélé, Y. K., Eyangoh, S., Verger, P., Leblanc, J. C., Le Bizec, B., & Krska, R. (2019). Regional sub-saharan Africa total diet study in benin, cameroon, mali and nigeria reveals the presence of 164 mycotoxins and other secondary metabolites in foods. *Toxins*, 11(1), 1–23. <https://doi.org/10.3390/toxins11010054>
- Jallow, E. A., Twumasi, P., Charles Mills-Robertson, F., & Dumevi, R. (2018). Assessment of aflatoxin-producing fungi strains and contamination levels of aflatoxin B1 in groundnut, maize, beans and rice. *Journal of Agricultural Science and Food Technology*, 4(4), 71–79. <http://pearlresearchjournals.org/journals/jasft/index.html>
- Kachapulula, P. W., Akello, J., Bandyopadhyay, R., & Cotty,

- P. J. (2017). Aspergillus section Flavi community structure in Zambia influences aflatoxin contamination of maize and groundnut. *International Journal of Food Microbiology*, 261, 49–56. <https://doi.org/10.1016/j.ijfoodmicro.2017.08.014>
- Kamala, A., Kimanya, M., De Meulenaer, B., Kolsteren, P., Jacxsens, L., Haesaert, G., Kilango, K., Magoha, H., Tiisekwa, B., & Lachat, C. (2018). Post-harvest interventions decrease aflatoxin and fumonisin contamination in maize and subsequent dietary exposure in Tanzanian infants: A cluster randomised-controlled trial. *World Mycotoxin Journal*, *In press*. <https://doi.org/10.3920/WMJ2017.2234>
- Kang'Ethe, E. K., Sirma, A. J., Murithi, G., Mburugu-Mosoti, C. K., Ouko, E. O., Korhonen, H. J., Nduhiu, G. J., Mungatu, J. K., Joutsjoki, V., Lindfors, E., & Ramo, S. (2017). Occurrence of mycotoxins in food, feed, and milk in two counties from different agro-ecological zones and with historical outbreak of aflatoxins and fumonisins poisonings in Kenya. *Food Quality and Safety*, 1(3), 161–169. <https://doi.org/10.1093/fqsafe/fyx018>
- Kebede, H., Liu, X., Jin, J., & Xing, F. (2020). Current status of major mycotoxins contamination in food and feed in Africa. *Food Control*, 110, 106975. <https://doi.org/10.1016/j.foodcont.2019.106975>
- Khaneghah, A. M., Ismail, E., Raeisi, S., & Fakhri, Y. (2018). Aflatoxins in cereals: State of the art. *Journal of Food Safety*, 38(6), 1–7. <https://doi.org/10.1111/jfs.12532>
- Kortei, N. K., Agyekum, A. A., Akuamo, F., Baffour, V. K., & Alidu, W. H. (2019). Risk assessment and exposure to levels of naturally occurring aflatoxins in some packaged cereals and cereal based foods consumed in Accra, Ghana. *Toxicology Reports*, 6, 34–41. <https://doi.org/10.1016/j.toxrep.2018.11.012>
- Kortei, N. K., Annan, T., Akonor, P. T., Richard, S. A., Annan, H. A., Kyei-Baffour, V., Akuamo, F., Akpaloo, P. G., & Esua-Amofo, P. (2021). The occurrence of aflatoxins and human health risk estimations in randomly obtained maize from some markets in Ghana. *Scientific Reports*, 11, 1–13. <https://doi.org/10.1038/s41598-021-83751-7>
- Kumari, J. W. P., Wijayarathne, L. K. W., Jayawardena, N. W. I. A., & Egodawatta, W. C. P. (2020). Quantitative and Qualitative Losses in Paddy, Maize and Greengram Stored under Household Conditions in Anuradhapura District of Sri Lanka. *Sri Lankan Journal of Agriculture and Ecosystems*, 2(1), 99–106. <https://doi.org/10.4038/sljae.v2i1.32>
- Kunz, B. M., Wanko, F., Kemmlin, S., Bahlmann, A., Rohn, S., & Maul, R. (2020). Development of a rapid multi-mycotoxin LC-MS/MS stable isotope dilution analysis for grain legumes and its application on 66 market samples. *Food Control*, 109, 106949. <https://doi.org/10.1016/j.foodcont.2019.106949>
- Likhayo, P., Bruce, A. Y., Tefera, T., & Mueke, J. (2018). Maize grain stored in hermetic bags: Effect of moisture and pest infestation on grain quality. *Journal of Food Quality*, 2515698, 1–9. <https://doi.org/10.1155/2018/2515698>
- Lorenzo, M., Sabrina, S., Gianpaola, P., Antonio, M., Miriam, H., & Giovanni, V. (2020). N2 controlled atmosphere reduces postharvest mycotoxins risk and pests attack on cereal grains. *Phytoparasitica*, 48(4), 555–565. <https://doi.org/10.1007/s12600-020-00818-3>
- Mahato, D. K., Lee, K. E., Kamle, M., Devi, S., Dewangan, K. N., Kumar, P., & Kang, S. G. (2019). Aflatoxins in Food and Feed: An Overview on Prevalence, Detection and Control Strategies. *Frontiers in Microbiology*, 10, 1–10. <https://doi.org/10.3389/fmicb.2019.02266>
- Mannaa, M., & Kim, K. D. (2017). Control Strategies for Deleterious Grain Fungi and Mycotoxin Production from Preharvest to Postharvest Stages of Cereal Crops : A Review. *Life Science and Natural Resources Research*, 25, 13–27. <https://www.researchgate.net/publication/323028050>
- Maringe, D. T., Chidewe, C., Benhura, M. A., Mvumi, B. M., Murashiki, T. C., Dembedza, M. P., Siziba, L., & Nyanga, L. K. (2016). Natural postharvest aflatoxin occurrence in food legumes in the smallholder farming sector of Zimbabwe. *Food Additives and Contaminants: Part B Surveillance*, 10, 1–7. <https://doi.org/10.1080/19393210.2016.1240245>
- Mezgebe, A. G., Terefe, Z. K., Bosh, T., Muchie, T. D., & Teklegiorgis, Y. (2016). Post-harvest losses and handling practices of durable and perishable crops produced in relation with food security of households in Ethiopia: secondary data analysis. *Journal of Stored Products and Postharvest Research*, 7(5), 45–52. <https://doi.org/10.5897/JSPPR2016.0205>
- Michael, B., Chris, O., Babu, N., Aisha, A., Salisu Sanusi, G., Gaya, S., Alabi, O., & Adobe, K. (2018). Towards a successful management of aflatoxin contamination in legume and cereal farming systems in northern Nigeria: A case study of the groundnut value chain. *African Journal of Agriculture and Food Security*, 6(7), 269–276. www.internationalscholarsjournals.org
- Mollay, C., Kassim, N., Stoltzfus, R., & Kimanya, M. (2021). Complementary feeding in Kongwa, Tanzania: Findings to inform a mycotoxin mitigation trial. *Maternal and Child Nutrition*, e13188, 1–10. <https://doi.org/10.1111/mcn.13188>
- Mom, M. P., Romero, S. M., Larumbe, A. G., Iannone, L., Comerio, R., Smersu, C. S. S., Simón, M., & Vaamonde, G. (2020). Microbiological quality, fungal diversity and aflatoxins contamination in carob flour (*Prosopis flexuosa*). *International Journal of Food Microbiology*, 326(108655). <https://doi.org/10.1016/j.ijfoodmicro.2020.108655>
- Muhie, O. A., & Bayisa, A. B. (2020). Is Aflatoxin a Threat to Human-Health in Ethiopia? A Systematic Review. *International Journal of Collaborative Research on Internal Medicine & Public Health*, 12(4), 1007–1015.
- Mukhtar, H., Farooq, Z., & Manzoor, M. (2016). Determination of aflatoxins in super kernel rice types consumed in different regions of Punjab, Pakistan. *Journal of Animal and Plant Sciences*, 26(2), 542–548.
- Mzungu, I., Hamisu, H., & Umar, K. (2018). Evaluation of Moulds Contamination of Cereals and Legumes Sold in Dutsinma Metropolis and their Aflatoxin Production Potential. *FUDMA Journal of Sciences*, 2(4), 94–98.

<https://doi.org/10.1088/1751-8113/44/8/085201>

Ncube, J., & Maphosa, M. (2020). Current state of knowledge on groundnut aflatoxins and their management from a plant breeding perspective: Lessons for Africa. *Scientific African*, 7, e00264. <https://doi.org/10.1016/j.sciaf.2020.e00264>

Neme, K., & Mohammed, A. (2017). Mycotoxin occurrence in grains and the role of postharvest management as a mitigation strategies. A review. *Food Control*, 78, 412–425. <https://doi.org/10.1016/j.foodcont.2017.03.012>

Nesci, A., Passone, M. A., Barra, P., Girardi, N., García, D., & Etcheverry, M. (2016). Prevention of aflatoxin contamination in stored grains using chemical strategies. *Current Opinion in Food Science*, 11, 56–60. <https://doi.org/10.1016/j.cofs.2016.09.010>

Njoroge, A. W., Baoua, I., & Baributsa, D. (2019). Postharvest Management Practices of Grains in the Eastern Region of Kenya. *Journal of Agricultural Science*, 11(3), 33–42. <https://doi.org/10.5539/jas.v11n3p33>

Njoroge, S. M. C. (2018). A critical review of aflatoxin contamination of peanuts in Malawi and Zambia: The past, present, and future. *Plant Disease*, 102, 2394–2406. <https://doi.org/10.1094/pdis-02-18-0266-fe>

Nleya, N., Adetunji, M. C., & Mwanza, M. (2018). Current status of mycotoxin contamination of food commodities in Zimbabwe. *Toxins*, 10(5), 1–12. <https://doi.org/10.3390/toxins10050089>

Obade, M. (2015). Exposure of children 4 to 6 months of age to aflatoxin in Kisumu County, Kenya. *African Journal of Food, Agriculture, Nutrition and Development*, 15(2), 9950–9963.

Ojiambo, P. S., Battilani, P., Cary, J. W., Blum, B. H., & Carbone, I. (2018). Cultural and genetic approaches to manage aflatoxin contamination: Recent insights provide opportunities for improved control. *Phytopathology*, 108(9), 1024–1037. <https://doi.org/10.1094/PHYTO-04-18-0134-RVW>

Ojuri, O. T., Ezekiel, C. N., Eskola, M. K., Šarkanj, B., Babalola, A. D., Sulyok, M., Hajšlová, J., Elliott, C. T., & Krska, R. (2019). Mycotoxin co-exposures in infants and young children consuming household- and industrially-processed complementary foods in Nigeria and risk management advice. *Food Control*, 98, 312–322. <https://doi.org/10.1016/j.foodcont.2018.11.049>

Opoku, N., Achaglinkame, M. A., & Amagloh, F. K. (2018). Aflatoxin content in cereal-legume blends on the Ghanaian market far exceeds the permissible limit. *Food Security*, 10(6), 1539–1545. <https://doi.org/10.1007/s12571-018-0849-5>

Ponzilacqua, B., Corassin, C. H., & Oliveira, C. A. F. (2018). Antifungal Activity and Detoxification of Aflatoxins by Plant Extracts: Potential for Food Applications. *The Open Food Science Journal*, 10(1), 24–32. <https://doi.org/10.2174/1874256401810010024>

Portell, X., Verheeecke-Vaessen, C., Torrelles-Ràfales, R., Medina, A., Otten, W., Magan, N., & García-Cela, E. (2020). Three-dimensional study of *f. Graminearum* colonisation of

stored wheat: Post-harvest growth patterns, dry matter losses and mycotoxin contamination. *Microorganisms*, 8, 1–18. <https://doi.org/10.3390/microorganisms8081170>

Prakash, J. (2016). Safety of Fermented Cereals and Legumes. In V. Prakash, O. Martín-Belloso, L. Keener, S. Astley, S. Braun, H. McMahon, & H. Lelieveld (Eds.), *Regulating Safety of Traditional and Ethnic Foods* (1st ed., pp. 283–310). Elsevier Inc. <https://doi.org/10.1016/B978-0-12-800605-4.00014-1>

Pratap, A., Mehandi, S., Pandey, V. R., Malviya, N., & Katiyar, P. K. (2016). Pre- and Post-harvest Management of Physical and Nutritional Quality of Pulses. In U. Singh, C. S. Praharaaj, S. S. Singh, & N. P. Singh (Eds.), *Biofortification of Food Crops* (pp. 421–431). Springer India. <https://doi.org/10.1007/978-81-322-2716-8>

Quellhorst, H. E., Njoroge, A., Venort, T., & Baributsa, D. (2020). Postharvest management of grains in Haiti and gender roles. *Sustainability*, 12, 1–13. <https://doi.org/10.3390/su12114608>

Ramirez, M. L., Cendoya, E., Nichea, M. J., Zachetti, V. G. L., & Chulze, S. N. (2018). Impact of toxigenic fungi and mycotoxins in chickpea: a review. *Current Opinion in Food Science*, 23, 32–37. <https://doi.org/10.1016/j.cofs.2018.05.003>

Sabuncuoğlu, S., Erkekoglu, P., Aydin, S., Şahin, G., & Kocer-Gumusel, B. (2015). The effects of season and gender on the serum aflatoxins and ochratoxin A levels of healthy adult subjects from the Central Anatolia Region, Turkey. *European Journal of Nutrition*, 54, 629–638. <https://doi.org/10.1007/s00394-014-0744-6>

Sarrocco, S., & Vannacci, G. (2018). Preharvest application of beneficial fungi as a strategy to prevent postharvest mycotoxin contamination: A review. *Crop Protection, In Press*. <https://doi.org/10.1016/j.cropro.2017.11.013>

Schmidt, M. (2018). *Thesis presented by* [University College Cork, Ireland]. http://intranet.coleurop.pl/library/repository/2014_thesis_peekles.pdf

Schmidt, M., Zannini, E., & Arendt, E. K. (2018). Recent advances in physical post-harvest treatments for shelf-life extension of cereal crops. *Foods*, 7, 1–22. <https://doi.org/10.3390/foods7040045>

Schmidt, M., Zannini, E., & Arendt, E. K. (2019). Screening of post-harvest decontamination methods for cereal grains and their impact on grain quality and technological performance. *European Food Research and Technology*, 245(5), 1061–1074. <https://doi.org/10.1007/s00217-018-3210-5>

Seetha, A., Munthali, W., Msere, H. W., Swai, E., Muzanila, Y., Sichone, E., Tsusaka, T. W., Rathore, A., & Okori, P. (2017). Occurrence of aflatoxins and its management in diverse cropping systems of central Tanzania. *Mycotoxin Research*, 33, 323–331. <https://doi.org/10.1007/s12550-017-0286-x>

Serdar, S. A., Tawila, M. M. El, Madkour, M. H., & Alrasheedi, A. A. (2020). Determination of Aflatoxins (AFs) in Different Food Samples: A Case Study from Jeddah, Saudi Arabia. *Met., Env. & Arid Land Agric. Sci.*, 29(1), 23–34.

<https://doi.org/10.4197/Met>.

Sirma, A. J., Lindahl, J. F., Makita, K., Senerwa, D., Mtimet, N., Kang'ethe, E. K., & Grace, D. (2018). The impacts of aflatoxin standards on health and nutrition in sub-Saharan Africa: The case of Kenya. *Global Food Security*, 18, 57–61. <https://doi.org/10.1016/j.gfs.2018.08.001>

Sulaiman, S. H., Jamaluddin, R., & Sabran, M. R. (2018). Association between urinary aflatoxin (AFM1) and dietary intake among adults in Hulu Langat District, Selangor, Malaysia. *Nutrients*, 10(4), 1–15. <https://doi.org/10.3390/nu10040460>

Sun, X. D., Su, P., & Shan, H. (2017). Mycotoxin Contamination of Rice in China. *Journal of Food Science*, 82(3), 573–584. <https://doi.org/10.1111/1750-3841.13631>

Tawila, M. El, Sadeq, S., Awad, A. A., Serdar, J., Madkour, M. H. F., & Deabes, M. M. (2020). Aflatoxins contamination of human food commodities collected from Jeddah markets, Saudi Arabia. *Open Access Macedonian Journal of Medical Sciences*, 8(E), 117–126. <https://doi.org/10.3889/oamjms.2020.4643>

Telles, A. C., Kupski, L., & Furlong, E. B. (2017). Phenolic compound in beans as protection against mycotoxins. *Food Chemistry*, 214, 293–299. <https://doi.org/10.1016/j.foodchem.2016.07.079>

Temba, M. C., Njobeh, P. B., & Kayitesi, E. (2017). Storage stability of maize-groundnut composite flours and an assessment of aflatoxin B1 and ochratoxin A contamination in flours and porridges. *Food Control*, 71, 178–186. <https://doi.org/10.1016/j.foodcont.2016.06.033>

Tibagonzeka, J. E., Akumu, G., Kiyimba, F., Atukwase, A., Wambete, J., Bbemba, J., & Muyonga, J. H. (2018). Post-Harvest Handling Practices and Losses for Legumes and Starchy Staples in Uganda. *Agricultural Sciences*, 09, 141–156. <https://doi.org/10.4236/as.2018.91011>

Tola, M., & Kebede, B. (2016). Occurrence, importance and control of mycotoxins: A review. *Cogent Food & Agriculture*, 2(1), 1–26. <https://doi.org/10.1080/23311932.2016.1191103>

Tuan, P. A., Kumar, R., Rehal, P. K., Toora, P. K., & Ayele, B. T. (2018). Molecular mechanisms underlying abscisic acid/gibberellin balance in the control of seed dormancy and germination in cereals. *Frontiers in Plant Science*, 9, 1–14. <https://doi.org/10.3389/fpls.2018.00668>

Tumukunde, E., Ma, G., Li, D., Yuan, J., Qin, L., & Wang, S. (2020). Current research and prevention of aflatoxins in China. *World Mycotoxin Journal*, 13(2), 121–138. <https://doi.org/10.3920/WMJ2019.2503>

Udomkun, P., Tirawattanawanich, C., Ilukor, J., Sridonpai, P., Njukwe, E., Nimbona, P., & Vanlauwe, B. (2019). Promoting the use of locally produced crops in making cereal-legume-based composite flours: An assessment of nutrient, antinutrient, mineral molar ratios, and aflatoxin content. *Food Chemistry*, 286, 651–658. <https://doi.org/10.1016/j.foodchem.2019.02.055>

Udomkun, P., Wiredu, A. N., Nagle, M., Müller, J., Vanlauwe, B., & Bandyopadhyay, R. (2017). Innovative technologies to manage aflatoxins in foods and feeds and the profitability of application – A review. *Food Control*, 76, 127–138. <https://doi.org/10.1016/j.foodcont.2017.01.008>

Valencia-Quintana, R., Milić, M., Jakšić, D., Klarić, M. Š., Tenorio-Arvide, M. G., Pérez-Flores, G. A., Bonassi, S., & Sánchez-Alarcón, J. (2020). Environment changes, aflatoxins, and health issues, a review. *International Journal of Environmental Research and Public Health*, 17, 1–10. <https://doi.org/10.3390/ijerph17217850>

Waliyar, F., Osiru, M., Ntare, B. R., Vijay Krishna Kumar, K., Sudini, H., Traore, A., & Diarra, B. (2015). Post-harvest management of aflatoxin contamination in groundnut. *World Mycotoxin Journal*, 8(2), 245–252. <https://doi.org/10.3920/WMJ2014.1766>

Wangia-Dixon, R. N., Quach, T. H. T., Song, X., Ombaka, J., Githanga, D. P., Anzala, O. A., & Wang, J. S. (2020). Determinants of aflatoxin exposures in Kenyan School-aged children. *International Journal of Environmental Health Research*. <https://doi.org/10.1080/09603123.2020.1854192>

Wanjari, K. R., Katheriya, I. J., & Obimbo, L. P. (2017). Diversity of Micro, Small and Medium Cereal Milling Enterprises in Nairobi County, Kenya and Levels of Aflatoxins in Their Milled Products. *World Journal of Nutrition and Health*, 5(2), 33–40. <https://doi.org/10.12691/jnh-5-2-2>

Ware, L. Y., Durand, N., Nikiema, P. A., Alter, P., Fontana, A., Montet, D., & Barro, N. (2017). Occurrence of mycotoxins in commercial infant formulas locally produced in Ouagadougou (Burkina Faso). *Food Control*, 73, 518–523. <https://doi.org/10.1016/j.foodcont.2016.08.047>



©2021 This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 International license viewed via <https://creativecommons.org/licenses/by/4.0/> which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is cited appropriately.