



EVALUATION OF ETHANOL LEAF EXTRACTS OF SIDA ACUTA AND CHROMOLAENA ODORATA ON INSECT PESTS OF CELOSIA ARGENTEA L. AND AMARANTHUS CRUENTUS L.

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ABSTRACT

A field experiment was carried out to evaluate the effect of leaf extracts of *Sida acuta* Burm. f. and *Chromolaena odorata* L. in controlling insect pests of *Celosia argentea* L. and *Amaranthus cruentus* L. The experiment was laid out in randomized complete block design and the treatments were replicated three times. Extracts obtained from the plants at 240 g/L of ethanol were applied as botanical insecticides. These extracts were examined along with a synthetic insecticide (Cypermethrin) including untreated control. Data were collected on number of *Cletus* spp. and *Hymenia recurvalis* per plant. Result shows that *C. argentea* sole treated with *S. acuta* and *C. odorata* was not infested by *Cletus* spp., and significantly (p<0.05) different from *A. cruentus* and *C. argentea* intercrop treated with *S. acuta* and *C. odorata* at 2 weeks after treatment. *C. argentea* treated with *S. acuta* had the least number of *H. recurvalis* (1.00) which was not significantly (p>0.05) different from Cypermethrin in both sole and intercrop at 4 WAT. *A. cruentus* and *C. argentea* planted sole or in intercrop treated with botanical and synthetic insecticides had lower numbers of *H. curvalis* which were significantly different from untreated control at 8 and 10 WAT. This study showed that *S. acuta* performed better than *C. odorata* against the target insect pests and therefore recommended for incorporation into Integrated Pest Management irrespective of whether sole or intercropping was adopted.

Keywords: Amaranthus, Celosia, Cletus, extracts, pests, Hymenia, vegetables

INTRODUCTION

Amaranthus spp. L. is an important vegetable in Nigeria, grown mainly for its highly nutritious leaves. Leaf amaranth is used as a steamed vegetable in soups and stews. The leaves, stem and head are reportedly high in protein (15 to 24% on a dry matter basis) (Directorate Plant Protection, 2010). Marin et al. (2008) listed Amaranthus as one of the crops that can respond to different ecological conditions and has become important for agro-food production and non-food sectors (biomass, biofuels, natural dyes, medicines). The leaves are very rich in proteins and micronutrients such as iron, calcium, zinc, vitamin C and vitamin A (Achigan-Dako et al., 2014). It has been reported that the amount of minerals such as calcium, magnesium, iron and zinc in wheat grain are 5.2-, 2.9-, 2.8and 1.3-fold lower than in amaranth seeds, respectively (Alvarez-Jubete et al., 2010). However, whole grains contain significant amounts of phytic acid, a well-known inhibitor of iron absorption and other minerals (Sanz-Penella et al., 2012). Amaranths can therefore, be used to reduce hunger and malnutrition, particularly in sub-Saharan Africa. This

necessitated the establishment of Biodiversity International's African leafy vegetables programme conducted in Botswana, Cameroon, Kenya, Senegal and Zimbabwe on growing, consumption, marketing and nutritional awareness of African leafy vegetables including amaranths (Gotor and Irungu, 2010). Amaranthus hybridus is consumed as a staple food in Africa and North America, Asia, the Caribbean and parts of Europe (Abbasi et al., 2013). Amaranthus leaves show significant energy value ranging from 27 to 53 kcal/100 g of fresh leaves and high nutrient value ranging from 4 to 6 g of protein, 0.2 to 0.6 g of fat, and 4 to 7 g of carbohydrates per 100 g of fresh leaves (Uusikua et al., 2010). As compared to lettuce, Amaranthus leaves contain 18 times more vitamin A, 13 times more vitamin C, 20 times more calcium and 7 times more iron (Srivastava, 2011). The folic acid in amaranth reduces the risk of neural defects in pregnant women and their new-borns (AVRDC, 2011).

Celosia argentea is one of the important leafy vegetable in south-western Nigeria. It is a vegetable of high economic value particularly in the dry season, as it provides a source of living for most rural vegetable farmers (Akinfasoye *et al.*, 2008). The

leaves and succulent stem are consumed as vegetable because it constitutes a cheap and rich nutrient source for the low income earners, and the seeds could also be processed into food items, supplement and additives. It also has some medicinal properties for example, in Kenya, the Mai Sai use the liquid extract from the leaves and flowers to bath a patient recovering from illness and also as an antidote for snake bites.

One of the limiting factors that affecting the productivity of amaranths is the range of insect pests (Aderolu et al., 2013). Borisade (2019) recorded a total of nine pests on amaranth from three insect orders namely, Ortoptera (62.5%), Coleoptera (12.5%) and Lepidoptera (25.0%). The insect pests that are responsible for the most economic damage to leaf amaranth in the Southwest Nigeria belong to Lepidoptera and Orthoptera orders (Borisade and Uwaidem, 2017). Along with Lepidoptera the other orders namely; Coleoptera, Hemiptera and Orthoptera are affect amaranths. Lepidoptera largely attack amaranths among the other order insect pests. The production of Amaranthus spp. is challenged by insect infestation including Amaranth stem weevils, Hypolixus truncatulus (F.), H. nubilosus (B.), Beeworm moth, Spoladea recurvalis (F.), leaf miner, Liriomyza huidobrensis B.), Aphid, Myzus persicae; plant bugs, Cletus sp. (Seni, 2018).

The use of synthetic insecticides in vegetable pest management particularly at unsafe levels causes pesticide residues and harm to the consumer. Despite the ban on use of DDT in the management of agricultural pests, Borisade *et al.* (2019) reported that the pesticide is still being used in the management of vegetable pests by subsistent farmers. Efforts should therefore be intensified by stakeholders on the need to find environmentally friendly alternatives to the use of inorganic pesticides in the control of pests of leaf vegetables. A sustainable alternative should be sought to the use of these synthetic insecticides.

The work therefore aims at evaluating the effects of ethanolic leaf extracts of common wire weed, *Sida acuta* Burm f. and siam weed, *Chromolaena odorata* on insect pests of *A. cruentus* and *C. argentea* planted sole or intercrop.

MATERIALS AND METHODS

The experiment was carried out during the rainy season of 2018 behind the Faculty of Agriculture, University of Ilorin, Ilorin, Nigeria. The study location falls within the southern Guinea Savannah agro-ecological zone of Nigeria between latitude 8°25'N and longitude 4°67'E. The rainfall ranges between 1200 and 1500 mm per annum. The climate includes rainy season which starts in April and ends in September while the dry season commences in October and ends in March. The dry spell is experienced in August of every year. The site has an average temperature of 35°C and is characterized by sandy loam soil (Babatunde *et al.* 2020).

The total land area was 21 m \times 43 m (903 m²). The nursery was situated beside the main plot. The land was manually

cleared, tilled and then pegged and divided into 12 blocks of 3 m \times 21 m from which each was sub-divided into 3 plots of 3 m \times 5 m and 1 m spacing between plots. The seeds of *A. cruentus* and *C. argentea* used were obtained from Agro Mall at Zango Area, Ilorin, Kwara State, Nigeria. The seeds were sown using broadcasting method and routine nursery management was carried out. The seedlings were transplanted at 5-leaf stage at a spacing of 1 m \times 1 m. The seedlings were randomly assigned to plots as *A. cruentus* sole and *C. argentea* sole, and *A. cruentus* and *C. argentea* intercrop.

Chromolaena odorata and S. acuta leaves were sourced from the University of Ilorin. These leaves were air dried for three (3) weeks and ground using electric blender, sieved by passing through 2 mm mesh to obtain a uniform particle size. The preparation of the extracts was done by maceration of 240 g/L of ethanol for 24 h separately and then filtered using muslin cloth. Each filtrate was made up to 2 L by adding distilled water and a 2 ml of liquid soap per litre of the solution was used as surfactant (Anjarwalla, 2016). The synthetic insecticide was prepared according to the manufacturer's recommendation (2 mL L-1 of water) while application of treatments commenced 2 weeks after transplanting (WAT) of seedlings according to Ezena et al. (2016) using hand sprayer and repeated at 2 weeks intervals up to 10 WAT. There were 12 treatment combinations that gave 48 experimental units. Ethanolic leaf extracts of S. acuta and C. odorata were used as botanical insecticides and (Cypermethrin) served as synthetic insecticide including an untreated control which was sprayed with water only. The experiment was laid out in a randomized complete block design with three replications.

Data were collected on target species such as *Cletus* spp. and *Hymenia recurvalis* plant after each spraying and continued at two weeks intervals up to 10 WAT. The insect populations from the different plots were sampled between 7 and 8.00 a.m. on each day of sampling. Number of insects/plant was sampled from inner parts of the plot. Data collected were subjected to two way analysis of variance using Genstat 17th edition, and significantly different means were compared using Fishers Least Significant Difference (LSD) test at 5 % level of significance.

RESULTS

Table 1 shows the effect of botanical and synthetic insecticides and *A. cruentus* and *C. argentea* planted sole or intercrop on the mean number of *Cletus* spp. The result shows that *C. argentea* sole treated with *S. acuta* and *C. odorata* was not infested by *Cletus* spp. and significantly (p<0.05) different from *A. cruentus* and *C. argentia* intercrop treated with *S. acuta* and *C. odorata* at 2 WAT. The *A. cruentus* sole and *A. cruentus* and *C. argentea* intercrop had mean numbers of *Cletus* spp. which were not significantly (p>0.05) different with Cypermethrin treated crops and the untreated control at 2 WAT. At 4 WAT, *A. cruentus* treated with *C. odorata* had mean number of *Cletus* spp. of 5.33 which was significantly (p<0.05) higher than the mean number of the insect in other treatments and the untreated control. The result shows that *A. cruentus* treated with *C. odorata*, and *A. cruentus* and *C. argentea* intercrop treated with *C. odorata* had mean number of *Cletus* spp. of 5.00 and 4.73 respectively, were not significantly (p>0.05) different from the mean number of *Cletus* spp. on untreated control (6.53) at 6 WAT.

Table 1 also shows that *A. cruentus* and *C. argentea* planted sole or intercrop treated with botanical and synthetic insecticides had mean numbers of *Cletus* spp. which were significantly (p<0.05) different from untreated control at 8 WAT. Similarly, *A. cruentus* and *C. argentea* planted sole or intercrop treated with botanical and synthetic insecticides had lower mean numbers of *Cletus* spp. which were significantly (p<0.05) different from untreated control except for *A. cruentus* treated with *C. odorata* at 10 WAT.

Table 1: Effect of insecticides and *Amaranthus cruentus* and *Celosia argentea* and sole and intercrop on number of *Cletus* spp.

	Cletus. Spp (WAP)								
Crop	Treatment	2	4	6	8	10			
A. cruentus	S. acuta	0.53ab	0.06 a	2.60ab	2.80abc	4.93ab			
	C. odorata	0.40ab	5.33c	5.00bc	4.60bcd	8.00bc			
	Cypermethrin	0.20ab	0.66ab	1.06a	1.80ab	2.20a			
C. argentea	S. acuta	0.00a	0.20a	5.06bc	2.20ab	3.00a			
	C. odorata	0.00a	2.06ab	3.06ab	5.46cd	5.66ab			
	Cypermethrin	0.13ab	1.46ab	0.53a	3.26abcd	3.16a			
A. cruentus + C. argentea	S. acuta	0.93b	1.40ab	2.80ab	5.66cd	4.06ab			
	C. odorata	0.93b	2.80b	4.73bc	6.20d	5.73ab			
	Cypermethrin	0.33ab	1.13ab	2.20ab	0.80a	2.73a			
	Control	0.13ab	2.66b	6.53c	9.33e	10.73c			
SEM		0.29	0.78	1.08	1.02	1.52			
LSD(0.05)		0.86	2.3	3.18	3.01	4.47			

Values in the same column followed by the same letter(s) are not significantly different at p = 0.05 according to Fisher's protected Least Significant Difference (LSD) test

Table 2 shows the effect of botanical and synthetic insecticides and *A. cruentus* and *C. argentea* planted sole or intercrop on the mean number of *Hymenia recurvalis*. There was no significant (p>0.05) difference in the mean number of *H. recurvalis* in the treated crops and untreated control at 2 WAT. It was observed that *C. argentea* treated with *S. acuta* had the least mean number of *H. recurvalis* (1.00) which was not significantly (p>0.05) different from Cypermethrin in both sole and intercrop at 4 WAT. *Amaranthus cruentus* sole, *C. argentea* sole and *A. cruentus* and *C. argentea* intercrop treated with *C. odorata* had no significant difference in the mean numbers of *H. recurvalis* and not significantly different from control at 6 WAP. *Amaranthus cruentus* and *C. argentea* intercrop treated with botanical and synthetic insecticides had lower mean numbers of *H. curvalis* which were significantly (p<0.05) different from untreated control at 8 and 10 WAP.

Crop	Hymenia recurvalis (WAT)							
	Treatments	2	4	6	8	10		
A. cruentus	S. acuta	1.33c	1.73ab	3.13ab	3.66abc	5.66abc		
	C. odorata	1.40c	4.46bc	4.46abcd	5.60cd	6.93bc		
	Cypermethrin	0.80abc	1.86ab	1.93a	2.26ab	2.53a		
C. argentea	S. acuta	1.33c	1.00a	3.66abc	5.13abcd	4.13abc		
	C. odorata	0.20ab	2.73abc	4.86bcd	5.00abcd	7.00c		
	Cypermethrin	0.66abc	0.66a	2.93ab	1.86a	2.97ab		
A. cruentus + C. argentea	S. acuta	0.80abc	1.53a	3.80abc	3.53abc	4.13abc		
	C. odorata	0.86abc	3.06abc	5.06bcd	5.53bcd	6.73bc		
	Cypermethrin	0.33abc	1.66a	3.73abc	3.46abc	4.80abc		
	Control	1.20bc	4.93c	6.66d	9.06e	12.13d		
SEM		0.36	0.94	0.91	1.12	1.35		
LSD(0.05)		1.07	2.75	2.67	3.29	3.97		

 Table 2: Effect of insecticides and Amaranthus cruentus and Celosia argentea sole and intercrop on number of Hymenia recurvalis

Values in the same column followed by the same letter(s) are not significantly different at p = 0.05 according to Fisher's protected Least Significant Difference (LSD) test

DISCUSSION

Researchers have adopted farming systems capable of improving the socio-economic sustainability in crop production and protection. The farming systems are known to play important role in management of insect pests which may include Cletus spp. and H. recurvalis. In this study, it was observed that S. acuta and C. odorata leaf extracts competed with Cypermethrin in reducing number of *Cletus* spp. and *H*. recurvalis irrespective of whether A. cruentus and C. argentea were planted sole or intercrop. It has been reported that C. odorata and neem extracts had significantly reduced number of aphids on cabbage than the conventional insecticide, Sunhalothrin (Ezena et al., 2016). The use of S. acuta extract performed better than C. odorata extract and the untreated control thereby corroborating its insecticidal activity as reported by other authors (Adeniyi et al., 2010; Narasimhon et al., 2014; Gadewad and Paedeshi, 2018). It was observed that number of insects varied depending on type of extract, crop and farming technique. However, the effect of plant extracts may have to be increased by repeated application due to quick breakdown of the active components. The efficacy of Cypermethrin against insects has been reported by several authors (Kwaifa *et al.*, 2012). Previous researchers studied the insecticidal activity of plant extracts against *Callosobruchus maculatus*, *Dysdercus cingulatus* (Gadewad and Pardeshi, 2017). Studies have revealed that *S acuta* and *C. odorata* contain phytochemicals that are effective in insect pest control (Adeniyi *et al.*, 2010; Govindarajan, 2010; Deepti *et al.*, 2013; Wahab and Akinterinwa, 2015).

CONCLUSION

The findings of this study revealed that ethanol extracts of *S. acuta* and *C. odorata* performed competitively with the synthetic insecticide (Cypermethrin) against the target insect

pests. In comparison to imported chemicals, these plant materials are available in Nigeria and easier to prepare and apply with no reported issues of environmental toxicity. The study also showed that the plant extracts were capable of reducing the number of *Cletus* spp. and *H. recurvalis* in *A. cruentus* and *C. argentea* planted sole or intercrop. The incorporation of the plant extracts in Integrated Pest Management is recommended against multiplication of insect pests in the field.

REFERENCES

Abbasi, A.M., Khan, M.A., Shah, M.H., Shah, M.M., Pervez, A. and Ahmad, M. (2013). Ethnobotanical appraisal and cultural values of medicinally important wild edible vegetables of Lesser Himalayas-Pakistan. *Journal of Ethnobiology and Ethnomedicine*, *9*: 66.

Achigan-Dako, E.G., Sogbohossou, O.E.D. and Maundu, P. (2014). Current knowledge on *Amaranthus* spp.: research avenues for improved nutritional value and yield in leafy amaranths in sub-Saharan Africa. *Euphytical* 197: 303-317.

Adeniyi, S.A., Orjiekwe, C.L., Ehiagbonare, J.E. and Arimah, B.D. (2010). Preliminary phytochemical analysis and insecticidal activity of ethanolic eaxtracts of four tropical plants (*Vernonia amygdalina, Sida acuta, Ocimum* gratissimum and Telfaria occidentalis) against bean weevil (*Acanthoscelides obtectus*). International Journal of Physical Sciences 5: 753-762.

Aderolu, I. A., Omooloye, A. A and Okelana, F. A. (2013). Occurrence, abundance and control of the major insect pests associated with amaranths in Ibadan, Nigeria. *Journal of Entomology, Ornithology & Herpetology, 2*: 112-120.

Anjarwalla, P., Belmain, S., Ofori, D.A., Sola, P., Jamnadass, R., Stevenson, P.C. (2016). *Handbook on Pesticidal Plants*. World Agroforestry Centre (ICRAF), Nairobi, Kenya.

AVRDC (2011). New variety releases expand market options for Tanzania's farmers. Fresh news from AVRDC-The World vegetable center pp: 1-3.

Babatunde, S. F., Ogunleye, S.T. and Solihu, A. A. (2020). Effects of leave and bark ash of *Azadirachta indica* extracts against insect pest of *Amaranthus hybridus* L. *International Journal of Agriculture and Environmental Research*. 6(3): 498-509.

Borisade, O.A., Awodele, S.O. and Uwaidem, Y.I. (2019). Insect pest profile of leaf amaranth (*Amaranthus hybridus* L.) and prevention of herbivory using oil-based extracts of *Allium* sativum L., *Xylopia aethiopica* Dunal and *Eucalyptus* globolus L. International Journa of Plant & Soil Science. 28(6): 1-9.

Deepti Malhotra, Amir Khan and Fouzia Ishaq (2013). Phytochemical screening and antibacterial effect of root extract of *Boerhavia diffusa L*. (Family Nyctaginaceae). *Journal of Applied and Natural Science*, 5(1), 221-225. <u>https://doi.org/10.31018/jans.v5i1.310</u>

Directorate Plant Protection (2010). Amaranthus production guidelines. Department of Agriculture, *Forestry and Fisheries*, South Africa. 24.

Ezena, G.N., Akotsen-Mensah, C. and Fening, K.O. (2016). Insecticidal potential of the invasive Siam weed *Chromolaena odorata* L. (Asteraceae) in the management of the major pests of cabbage and their natural enemies in southern Ghana. *Advances in Crop Science and Technology* 4: 230.

Gadewad, M. G. and Paedeshi, A. (2017). Insecticidal effect of chrysanthemum indicum against red cotton bug, *Dysdercus cingulatus* Fab. *International Journal of Recent Science Research* 8(12): 22380-22383.

Gadewad, M. G. and Paedeshi, A. (2018). Bioinsecticidal effect of Sida acuta plant extract against red cotton bug, *Dysdercus cingulatus Fab. International Journal of Zoology Studies* 3(1): 177-181.

Gotor, E. and Irungu, C. (2010). The impact of Biodiversity International's African leafy vegetables programme in Kenya. *Impact Assess Project Apprais* 28(1): 41-55.

Govindarajan, M. (2010). Larvicidal and repellent activities of *Sida acuta* Burm. F. (Family: Malvaceae) against three important vector mosquitoes. *Asian Pacific Journal of Tropical Medicine* **3**(9): 691–695.

Kwaifa, N.M., Ibrahim, N.D., Dike, M.C. and Abubakar, M.G. (2012). Bioefficacy of neem and mahogany extracts for control of cowpea flower thrips, *Megalurothrips sjostedti* Trybom (Thysanoptera: Thripidae) and grasshopper, *Ailopus simulatrix* Walker (Orthoptera: Pyrgomorphidae) in Sudan Savanna, Nigeria. *Nigerian Journal of Entomology* 29: 31-42.

Marin, D.I., Babeanu, N.E. and Popa, O. (2008). *Biodiversity conservation by promoting alternative crops.* Proceeding of the International Symposium on New Research in Biotechnology, Series F, USAMV Bucharest.

Narasimhar, A., Kadarkarai, M., Pari, M., Thiyagarajan, N., Arjunan, N., Kandasamy, K., Jiang-Shiou, H., Donald, R., Barnard, Hui, W.R. and Chandrasekar, A.A. (2014). Effect of *Sida acuta* and *Vetiveriazi zanioides* against the malarial vector, Anopheles stephensi and malarial parasite, *Plasmodium beraheli*. *International Journal of Pure and Applied Zoology* 2(1): 51-60.

Nneji, E.T., Uddin II, R.O. and Musa, A.K. (2020). Effects of *Boerhavia diffusa* L. nom. cons. and *Chromolaena odorata* (L.) R.M. King & H. Rob. extracts on some field insect pests of okra (*Abelmoschus esculentus* (L.) Moench.) *Acta Agriculturae Slovenica.* 115(1): 133-139.

Sanz-Penella, J.M., Laparra, J.M., Sanz, Y. and Haros, M. (2012). Bread supplemented with amaranth (*Amaranthus*

Uusikua, N. P., A. Oelofsea, K. G. Duodub, M. J. Besterc & M. Faberd. (2010). Nutritional value of leafy vegetables of sub-Saharan Africa and their potential contribution to human health: A review. In: *Journal of Food Composition and Analysis 23*: 499–509.

Wahab, O. M. and Akinterinwa, O. (2015). Phytochemical evaluation of the insecticidal potential of some ethnobotanicals against bean weevil (*Acanthscelides obtectus*). *African Journal of Basic and Applied Sciences* **7**(6): 298–302

cruentus): effect of phytates on in-vitro iron absorption. *Plant Foods Human Nutrition* 67(1): 50-56.

Seni, A. (2018). Insect pests of amaranthus and their management. *International Journal of Environment, Agriculture and Biotechnology (IJEAB)* 3(3): 1100-1103.

Srivastava, R. (2011). Nutritional quality of some cultivated and wild species of *Amaranthus L. International Journal of Phamaceutical Science and Research* 2: 3152–3156.



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